

Protein Purification

1) Salt fractionation

proteins are poorly soluble in pure water

Salt in: increase salt to increase solubility

by increasing electrostatic and H-bonds

Salt out: Very high concentration of ions, so less water hydrates proteins \rightarrow proteins aggregates and form hydrophobic interaction \rightarrow precipitation

2) Dialysis

separation of protein sample through a semi-permeable membrane

Gel filtration

(Size exclusion chromatography)

(Molecular sieve chromatography)

separate according to size

using porous beads

Small proteins are included into the beads and precipitate later

Larger proteins are excluded from the beads and precipitate faster

3) Chromatography \rightarrow uses a stationary phase of beads

Ion Exchange Chromatography

separation according to charge

$\oplus \rightarrow$ Basic $\rightarrow \uparrow$ pI

$\ominus \rightarrow$ acidic $\rightarrow \downarrow$ pI

Anionic exchange } Cationic exchange

use \oplus beads

use \ominus beads

Elution order

Elution order

\oplus then \ominus

\ominus then \oplus

Affinity chromatography

use molecules (chemical or antibodies) that bind specific protein with high affinity

☆ Example:

Concanavalin A purified by Glucose

SDS-PAGE

SDS is used to:

1) denature and solubilize proteins

2) Give proteins uniform \ominus charges

Reducing agent

\rightarrow β -mercaptoethanol

dithiothreitol

\rightarrow Reduce disulfide bonds

☆ It separates according to size

Small = faster, large = slower

4) Gel electrophoresis \rightarrow Gel = polyacrylamide

Isoelectric focusing

☆ uses a gel with pH gradient

So separate according to charge

☆ a protein stops where $\text{pH} = \text{pI}$



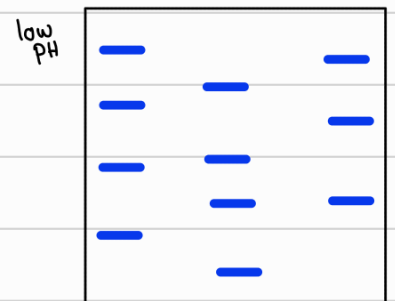
2D - PAGE

separate according to

charge and size

Isoelectric focusing

SDS PAGE



5) Immunoblotting (western blotting)

SDS-PAGE then add antibodies

Primary antibody \rightarrow bind the needed protein

Secondary antibody \rightarrow bind the Primary antibody
and has an enzyme on its E

6) ELISA \rightarrow detect protein and its amount more rapid, convenient and sensitive

Capture antibody \rightarrow bind the needed protein
 \hookrightarrow bound to a solid surface

Secondary antibody \rightarrow bind the needed protein

Protein sequencing

1) Edman degradation \rightarrow uses phenyl isothiocyanate (PITC) which reacts with the N-terminus
 \rightarrow can be used only for short fragments (50 amino acids)

2) Cleavage method

\hookrightarrow used for the whole protein

A) Chemical digestion

\hookrightarrow uses Cyanogen bromide (CNBr) \rightarrow cleave on C-terminus of Met

B) Endopeptidase

☆ Trypsin \rightarrow cleave on C-terminus of Arg, Lys

☆ Chymotrypsin \rightarrow cleave on C-terminus of Phe, Tyr, Trp

☆ Elastase \rightarrow cleave on C-terminus of Ala, Gly, Ser, Val

☆ Pepsin \rightarrow cleave on N-terminus of Leu, Phe, Tyr, Trp

\rightarrow Not next to Proline

C) Exopeptidase

\hookrightarrow Amino peptidase \rightarrow cleave on the N-terminus

\hookrightarrow Carboxy peptidase \rightarrow cleave on the C-terminus

\rightarrow ends

Determination of 3D structure

1) Crystallography

\hookrightarrow involves removing water

2) Nuclear magnetic Resonance (NMR)

spectroscopy

\hookrightarrow without removing water (in solution)

\hookrightarrow detect the dynamic and conformational changes